



TOYOHASHI UNIVERSITY of TECHNOLOGY

Tempaku-cho, Toyohashi, Aichi, 441-8580 Japan

PHONE: +81-532-44-6577 FAX: +81-532-44-6557

E-mail: press@office.tut.ac.jp

PRESS RELEASE

Source: Toyohashi University of Technology, Japan, Committee for Public Relations

Title: Micro fingers for arranging single cells

Subtitle: Development of hollow microprobe array for handling single cells in a parallel layout

Summary:

Toyohashi Tech researchers have developed a cell manipulation tool to trap and release single cells in a parallel arrangement in open-top microwells. Using microfabrication techniques, the researchers developed a hollow microprobe array with designed parameters. Single cells were trapped by suction and released by a flow generated through the probes. In the future, this tool will enable the reconstruction of microenvironments of stem cells, which can be employed to investigate stem cells for regenerative medicine.

Full text:

Functional analysis of a cell, which is the fundamental unit of life, is important for gaining new insights into medical and pharmaceutical fields. For efficiently studying cell functions, it is essential to reconstruct cellular microenvironments by parallel manipulation of single cells. Various cell manipulation techniques including fluidic, optical, and electrical techniques have been developed.

However, all these techniques lack flexibility with respect to changes in the cellular types, number, and places. In addition, the manipulations, which have been conducted in enclosed environments such as micro-channels, have limited access to the cells. It would be ideal if the working of a manipulation tool is analogous to grasping and transporting cells by one's fingers.

Now, researchers at Toyohashi Tech, at the Department of Mechanical Engineering, have developed a novel cell-manipulation tool that can trap and release single cells in a parallel arrangement in open-top microwells. The researchers fabricated an array of hollow microprobes to improve throughput and achieve flexibility in single-cell manipulation. The microprobes work like micro fingers picking up human cells. Single cells were trapped by suction and released by a flow generated through microchannels along the center of the probes.

"Parallel and versatile cell manipulation tools are essential for biomedical innovation", said Assistant Professor Moeto Nagai. "Microfabrication technologies offer massively parallel microstructures close to a human cell in size," he said.

"We fabricated an array of hollow microprobes with designed diameters, heights, and numbers from



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a silicon substrate using microfabrication techniques. Single cells were trapped on the tips of the probes using a suction flow. The cells were then released and placed in an array of microwells," Nagai said.

The research team developed a design principle for probes for minimally invasive single-cell manipulation. They conducted a cell aspiration experiment with a glass pipette and modeled a cell, thereby gaining information for designing hollow stepped probes. Based on the findings, the researchers designed and fabricated optimal probes. After a process of trial and error, the cells were placed and cultured in microwells with the probes, and the cells continued to maintain cell activity.

The proposed manipulation system makes it possible to place cells in a microwell array and observe the adherence, spreading, culture, and death of the cells. This system has the potential to be used as a tool for three-dimensional hetero cellular assays. In the future, this system will be further developed to reconstruct microenvironments of stem cells outside a living body, which would aid studies on stem cells for regenerative medicine.

Reference:

Nagai, M., Oohara, K., Kato, K., Kawashima, T., and Shibata, T. (2015). Development and characterization of hollow microprobe array as a potential tool for versatile and massively parallel manipulation of single cells, *Biomedical Microdevices*, 17(2):41. doi: 10.1007/s10544-015-9943-z.

Further information

Toyohashi University of Technology

1-1 Hibarigaoka, Tempaku, Toyohashi, Aichi Prefecture, 441-8580, JAPAN

Inquiries: Committee for Public Relations

E-mail: press@office.tut.ac.jp

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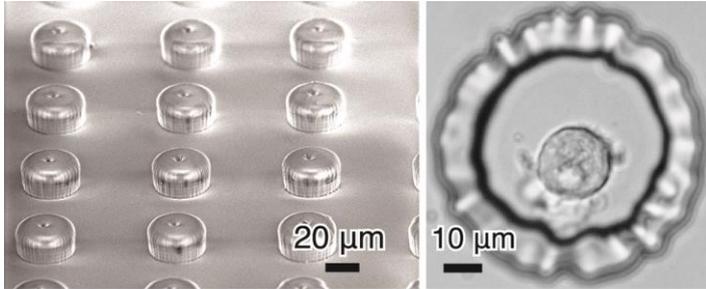


Figure 1:

Caption: The developed microprobe array and a single cell placed in a microwell.

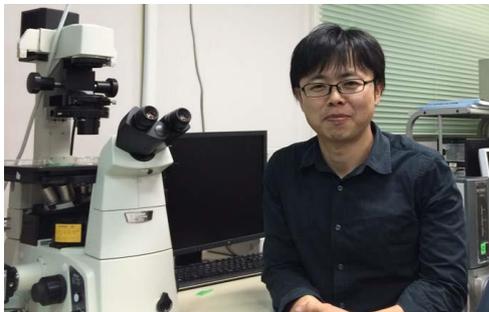


Figure 2:

Caption: Assistant Professor Moeto Nagai.

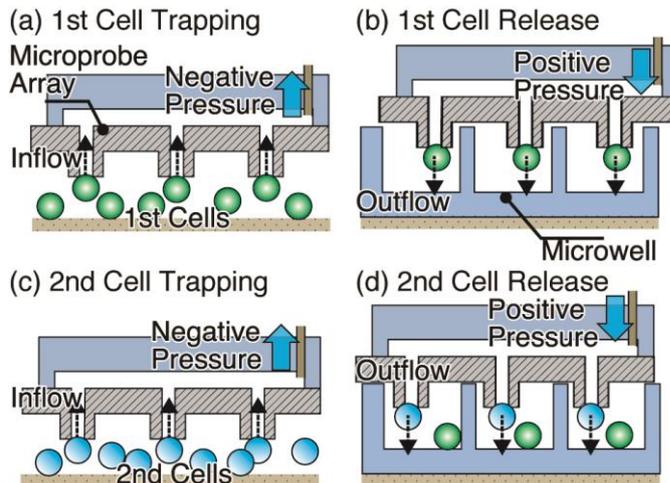


Figure 3:

Caption: Principle of single-cell manipulation in an array-based format.

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